



## ANNEXIN V-FITC STAINING PROTOCOL

Cat. No. CTG-AP0005

1 Kit (100 tests)

Annexin V-FITC Apoptosis Detection Kit is used to quantitatively determine the percentage of cells within a population that are actively undergoing apoptosis. It relies on the property of cells to lose membrane asymmetry in the early phases of apoptosis. In apoptotic cells, the membrane phospholipid phosphatidylserine (PS) is translocated from the inner leaflet of the plasma membrane to the outer leaflet, thereby exposing PS to the external environment. Annexin V is a calcium-dependent phospholipid-binding protein that has a high affinity for PS, and is useful for identifying apoptotic cells with exposed PS. Propidium Iodide (PI) is a standard flow cytometric viability probe and is used to distinguish viable from nonviable cells. Viable cells with intact membranes exclude PI, whereas the membranes of dead and damaged cells are permeable to PI. Cells that stain positive for FITC Annexin V and negative for PI are undergoing apoptosis. Cells that stain positive for both FITC Annexin V and PI are either in the end stage of apoptosis, are undergoing necrosis, or are already dead. Cells that stain negative for both FITC Annexin V and PI are alive and not undergoing measurable apoptosis.

### Reagents Preparation

1. FITC Annexin V: Use 5  $\mu$ l per test.
2. Propidium Iodide (PI) is a convenient, ready-to-use nucleic acid dye. Use 5  $\mu$ l per test.
3. 10X Annexin V Binding Buffer: 0.1 M HEPES/NaOH (pH 7.4), 1.4 M NaCl, 25 mM  $\text{CaCl}_2$ . For a 1X working solution, dilute 1 part of the 10X Annexin V Binding Buffer to 9 parts of distilled water.

### Staining

- 1) Wash cells twice with cold PBS and then resuspend cells in 1X Binding Buffer at a concentration of  $1 \times 10^6$  cells/ml.
- 2) Transfer 100  $\mu$ l of the solution ( $1 \times 10^5$  cells) to a 5 ml culture tube.
- 3) Add 5  $\mu$ l of FITC Annexin V and 5  $\mu$ l PI.
- 4) Gently vortex the cells and incubate for 15 min at RT (25°C) in the dark.
- 5) Add 400  $\mu$ l of 1X Binding Buffer to each tube. Analyze by flow cytometry within 1 hr.

The following controls are used to set up compensation and quadrants:

- Unstained cells.
- Cells stained with FITC Annexin V (no PI).
- Cells stained with PI (no FITC Annexin V).

### References

1. Schmid I, Krall WJ, Uittenbogaart CH, Braun J, Giorgi JV. Cytometry. 1992; 13(2):204-208.
2. Vermes I, Haanen C, Steffens-Nakken H, Reutelingsperger C. J Immunol Methods. 1995; 184(1):39-51.
3. Andree HA, Reutelingsperger CP, Hauptmann R, Hemker HC, Hermens WT, Willems GM. J Biol Chem. 1990; 265(9):4923-4928.
4. Casciola-Rosen L, Rosen A, Petri M, Schlissel M. Proc Natl Acad Sci U S A. 1996; 93(4):1624-1629.
5. Homburg CH, de Haas M, von dem Borne AE, Verhoeven AJ, Reutelingsperger CP, Roos D. Blood. 1995; 85(2):532-540.



CELLTECHGENLLC

# Celltechgen LLC

For Research Only

---

6. Koopman G, Reutelingsperger CP, Kuijten GA, Keehnen RM, Pals ST, van Oers MH. Blood. 1994; 84(5):1415-1420.
7. Martin SJ, Reutelingsperger CP, McGahon AJ, et al. J Exp Med. 1995; 182(5):1545-1556.
8. O'Brien MC, Bolton WE. Cytometry. 1995; 19(3):243-255.